

Highly sensitive targeted and non-targeted HPLC-MS analysis of PFAS

Per- and polyfluoroalkyl substances (PFAS) are persistent environmental pollutants known for their long lifetime and mobility. Their stability leads to accumulation in groundwater and soil, with proven harmful health effects. To mitigate environmental pollution, several PFAS compounds, such as perfluorooctanoic acid (PFOA), perfluorohexanesulfonic acid (PFHxS), and long-chain perfluoroalkyl carboxylic acids (C9-C14), are now regulated. This technical note presents a method for identifying and quantifying PFAS using a YMC-Triart C18 HPLC column. This column, based on a robust hybrid silica particle, offers enhanced separation of isomers and improved analytical performance when coupled with mass spectrometry.





Table 1: Chromatographic conditions.

Column:	YMC-Triart C18 (12 nm, 3 μm) 100 x 2.1 mm ID
Part No.:	IA12S03-10Q1P1H
Eluent:	A) water/methanol (95/5) + 2 mM ammonium acetate
	B) water/methanol (5/95) + 2 mM ammonium acetate
Gradient:	15–70 %B (0–2 min), 70–90 %B (2–5 min), 90–100 %B (5–10 min), 100 %B (10–15 min),
	15 %B (15.1–22 min)
Flow rate:	0.3 mL/min
Temperature:	40°C
Injection:	2μL
Detection:	ESI-MS negative mode
Sample:	PFAS standards (5, 10, 25, 50, 75, 100 ng/mL) from Fluka, Sigma, Dr. Ehrenstorfer, Apollo Scientific, soil extract sample (Brilon-Scharfenberg)

Table 2: MS source parameters.

Gas Temp:	150 °C
Gas Flow:	16 L/min
Nebulizer pressure:	35 psig
Sheath gas temperature:	380 °C
Sheath gas flow:	12 L/min
Fragmentor voltage:	380 V
Capillary voltage:	3000 V
Nozzle voltage:	300 V

Table 3: Substances included in the standard mixture.

Name	Abbreviation	Mass (Da)	m/z	Retention time (s)
Perfluorobutanoic acid	PFBA	213.9865	212.9792	291
Perfluoropentanoic acid	PFPeA	263.9833	262.9760	359
Perfluorohexanoic acid	PFHxA	313.9801	312.9728	398
Perfluoroheptanoic acid	PFHpA	363.9769	362.9696	432
Perfluorooctanoic acid	PFOA	413.9737	412.9664	469
Perfluorononanoic acid	PFNA	463.9705	462.9632	505
Perfluorodecanoic acid	PFDA	513.9673	512.9600	538
Perfluorobutanesulfonic acid	PFBS	299.9503	298.9430	366
Perfluorohexanesulfonic acid	PFHxS	399.9439	398.9366	435
Perfluorooctanesulfonic acid	PFOS	499.9375	498.9302	503
6:2 Fluorotelomer sulfonic acid	FTSA	427.9752	426.9679	467





Figure 1: Extracted ion chromatograms of (a) perfluoroalkyl sulfonic acids (PFSAs), (b) perfluoroalkyl carboxylic acids (PFCAs) and (c) perfluoroalkyl carboxylic acids (PFCAs) with loss of CO_2 during ionisation obtained with +/-120 seconds and +/-15 ppm at a concentration of 75 ng/ml each.



PFAS standards (Table 3) were analysed at concentrations ranging from 5–100 ng/mL. The standards could be well resolved by the YMC-Triart column (Figure 1). Notably, partial separation of PFOS and PFHxS isomers within 10 seconds facilitated fragmentation pattern analysis for structural elucidation.

expected (Figure 1b). Additionally, CO_2 -loss ions exhibited remarkably higher intensities than the primary molecular ions (Figure 1c). Calibration curves (Figure 2) demonstrated a linear quantification range, and carry-over tests confirmed no residual analytes after injections of 100 ng/mL. These results highlight the YMC-Triart C18 column's suitability for robust PFAS analysis.

For perfluoroalkyl carboxylic acids (PFCAs), higher peak intensities were observed for longer carbon chains as



Figure 2: Calibration curves of (a) perfluoroalkyl sulfonic acids (PFSAs), (b) perfluorinated carboxylic acids (PFCAs) and (c) perfluorinated carboxylic acids (PFCAs) with loss of CO₂ during ionisation at concentrations of 5-100 ng/mL.



Non-targeted analysis in a soil extract

To evaluate performance in complex matrices, a soil extract from a contaminated site (Brilon-Scharfenberg, Germany; from the scientific publication of Zweigle et al. [1]) was analysed. Total ion chromatograms (TICs) from triplicate measurements (Figure 3) revealed consistent profiles, despite the high matrix complexity. Table 4 lists preidentified PFAS clusters detected in the analysis (originally identified in Zweigle et al. [1] and shared by courtesy).



Figure 3: Total ion chromatograms (TIC) of the soil sample measured in triplicate.



Table 4: Substances identified in the soil extract by Zweigle et al. [1], using Kendrick mass defect analysis and matching of CF_2 -distances in the fragmentation spectra of prioritised chromatographic peaks.

Formula	m/z	Cluster Group	Formula	m/z	Cluster Group
$F(CF_2)_7SO_3$	448.93286		CF ₃ 0(CF ₂) ₈ SO ₃	564.921359	
F(CF ₂) ₈ SO ₃	498.92966		CF ₃ 0(CF ₂) ₉ SO ₃	614.918159	
F(CF ₂) ₉ SO ₃	548.92647		CF ₃ 0(CF ₂) ₁₀ SO ₃	664.914959	
F(CF ₂) ₁₀ SO ₃	598.92328		CF ₃ 0(CF ₂) ₁₁ SO ₃	714.911759	
F(CF ₂) ₁₁ SO ₃	648.92008	$\Gamma(U\Gamma_2)_n SU_3$	CF ₃ O(CF ₂) ₁₂ SO ₃	764.908559	$CF_3O(CF_2)_nSO_3$
F(CF ₂) ₁₂ SO ₃	698.9168		CF ₃ O(CF ₂) ₁₃ SO ₃	814.905359	
F(CF ₂) ₁₃ SO ₃	748.91369		CF ₃ O(CF ₂) ₁₄ SO ₃	864.902159	
$F(CF_2)_{14}SO_3$	798.9105		CF ₃ O(CF ₂) ₁₅ SO ₃	914.898959	
F(CF ₂) ₁₅ SO ₃	848.90731		CF ₃ OC ₂ F ₂ (CF ₂) ₆ SO ₃	526.924553	
SF ₅ (CF ₂) ₆ SO ₃	506.901728		CF ₃ OC ₂ F ₂ (CF ₂) ₇ SO ₃	576.921353	
SF ₅ (CF ₂) ₇ SO ₃	556.898528		CF ₃ OC ₂ F ₂ (CF ₂) ₈ SO ₃	626.918153	
SF ₅ (CF ₂) ₈ SO ₃	606.895328		CF ₃ OC ₂ F ₂ (CF ₂) ₉ SO ₃	676.914953	
SF ₅ (CF ₂) ₉ SO ₃	656.892128	SF ₅ (CF ₂) ₀ SO ₃	CF ₃ OC ₂ F ₂ (CF ₂) ₁₀ SO ₃	726.911753	$CF_3OC_2F_2(CF_2)_nSO_3$
SF ₅ (CF ₂) ₁₀ SO ₃	706.888928	0 2.11 0	CF ₃ OC ₂ F ₂ (CF ₂) ₁₁ SO ₃	776.908553	
SF ₅ (CF ₂) ₁₁ SO ₃	756.885728		CF ₃ OC ₂ F ₂ (CF ₂) ₁₂ SO ₃	826.905353	
SF ₅ (CF ₂) ₁₂ SO ₃	806.882528		CF ₃ OC ₂ F ₂ (CF ₂) ₁₃ SO ₃	876.902153	
CF(CF ₂) ₆ SO ₃	460.932838		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₄ SO ₃	422.936031	
$CF(CF_2)_7SO_3$	510.929638		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₆ SO ₃	522.929631	
CF(CF ₂) ₈ SO ₃	560.926438		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₇ SO ₃	572.926431	
CF(CF ₂) ₉ SO ₃	610.923238		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₈ SO ₃	622.923231	
$CF(CF_2)_{10}SO_3$	660.920038	CF(CF ₂) _n SO ₃	FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₉ SO ₃	672.920031	$FC_2F_2C_2F_2(CF_2)_nSO_3$
CF(CF ₂) ₁₁ SO ₃	710.916838		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₁₀ SO ₃	722.916831	
CF(CF ₂) ₁₂ SO ₃	760.913638		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₁₁ SO ₃	772.913631	
CF(CF ₂) ₁₃ SO ₃	810.910438		FC ₂ F ₂ C ₂ F ₂ (CF ₂) ₁₂ SO ₃	822.910431	
$CF(CF_2)_{14}SO_3$	860.907238]			





Figure 4: Extracted ion chromatograms of (a) PFAS group $F(CF_2)_n SO_3$, (b) PFAS group $SF_5(CF_2)_n SO_3$, (c) PFAS group $CF(CF_2)_n SO_3$, (d) PFAS group $CF_3 O(CF_2)_n SO_3$, (e) PFAS group $CF_3 OC_2 F_2 (CF_2)_n SO_3$ and (f) PFAS group $FC_2 F_2 C_2 F_2 (CF_2)_n SO_3$.





Figure 4: Extracted ion chromatograms of (a) PFAS group $F(CF_2)_n SO_3$, (b) PFAS group $SF_5(CF_2)_n SO_3$, (c) PFAS group $CF(CF_2)_n SO_3$, (d) PFAS group $CF_3O(CF_2)_n SO_3$, (e) PFAS group $CF_3OC_2F_2(CF_2)_n SO_3$ and (f) PFAS group $FC_2F_2C_2F_2(CF_2)_n SO_3$ continued.

The YMC-Triart C18 column achieved effective separation of PFAS (Figure 4), including improved partial resolution of isomers compared to Zweigle et al. [1]. For instance, $SF_5(CF_2)_9SO_3$ isomers were separated (Figure 5), enabling the acquisition of distinct fragmentation spectra (Figure 6). Ion traces at 575 s and 600 s revealed differences in fragmentation patterns, indicating structural variations. Identifiable fragments are listed in Table 5. The larger fragments in longer retention indicate an exclusively linear isomer. Specifically, the shorter retention time and the smaller fragments suggest a branched isomer or a differently positioned SF₅-group.



Figure 5: Ion trace correspondent to $SF_5(CF_2)_9SO_3^-$ with m/z 656.8927 +/-20 ppm.





Figure 6: Comparison of the averaged fragmentation patterns of the ion trace $SF_5(CF_2)_gSO_3^-$ at 575 +/-15 seconds (grey) and at 600 +/-15 seconds (green).

Table 5: Identifiable fragments of the averaged fragmentation patterns of the ion trace $SF_5(CF_2)_3SO_3^{-1}$ at 575 and 600 seconds.

575 s		600 s		
Fragment	m/z	Fragment	m/z	
SF ₅ -	79.9592 Da	$(CF_2)_4 SO_3^-$	279.9440 Da	
$(CF_2)_3SO_3^{-1}$	229.9472 Da	(CF ₂) ₆ SO ₃ ⁻	379.9377 Da	
$(CF_{2})_{6}SO_{3}^{-}$	379.9377 Da	(CF ₂) ₈ SO ₃ -	479.9313 Da	

Conclusion

The YMC-Triart C18 column demonstrates robust performance for PFAS analysis. It delivers:

- Efficient separation of key PFAS standards with linear quantification.
- **Reproducible results** in complex matrices, such as soil extracts.
- Improved isomer separation, enabling structural elucidation through distinct fragmentation patterns.

The column provides a reliable and enhanced approach to targeted and non-targeted PFAS analysis, supporting environmental monitoring and regulatory compliance.

* Application data by courtesy of Ricardo Cunha, Institut für Umwelt & Energie, Technik & Analytik e. V. (IUTA) Duisburg, Germany, Boris Bugsel and Jonathan Zweigle.

References:

[1] Environ. Sci. Technol. 2023, 57, 6647–6655.